

FGF23 & Klotho

Author:

Justine Bacchetta, MD, PhD

Centre de Référence des Maladies Rénales Rares
Service de Néphrologie, Rhumatologie et Dermatologie Pédiatriques
Hôpital Femme Mère Enfant, Bron, France

CONTENT

KEY POINTS

	Introduction	5
1	FGF23: structure and biochemical properties	5
2	Animal models and FGF23	5
3	Klotho: structural properties, overview on its role in physiology and in mineral metabolism	6
4	FGF23 in human physiology	7
5	FGF23 and Klotho in genetic human diseases	9
6	FGF23 and Klotho deregulation during chronic kidney disease	10
7	FGF23, Klotho and the cardiovascular system	12
8	FGF23, Klotho and global health	14
9	Different assays for measuring circulating FGF23 and Klotho	14
9.1	FGF23	14
9.2	Klotho	15
10	When should we measure FGF23 in clinical practice?	16
11	Hot topics in research	16
12	Conclusion	18
13	References	18

KEY POINTS

Fibroblast Growth Factor 23 (FGF23) is a protein synthesized by osteocytes that has a key role in the 'bone-parathyroid-kidney' axis and the regulation of phosphate/calcium metabolism. FGF23 has three main effects: hypophosphatemia (through an inhibition of phosphate reabsorption in the proximal tubule), decreased PTH levels and decreased $1,25(\text{OH})_2$ Vitamine D levels (through an inhibition of 1α hydroxylase and an activation of 24 hydroxylase activity in the kidney). Off-targets of FGF23 have also been demonstrated, notably on cardiomyocytes and monocytes. The links between iron metabolism and FGF23 on one hand, and between FGF23 and bone mineralization on the other hand, seem also of importance, although still under investigation.

The single-pass transmembrane Klotho protein, an anti-aging protein, is required *in vivo* for FGF23-mediated receptor activation, at least for its renal effects. However, Klotho also has mineral effects by it-self: it can function as an enzyme modifying the sugar chains of TRPV5 in the distal tubule, leading to increased calcium reabsorption, and it can also directly regulate PTH synthesis. Through alternative splicing or ectodomain shedding of the transmembrane protein, the Klotho protein exists both as a secreted (α -Klotho) and a membrane protein whose extracellular domain could be shed from the cell surface and released into the circulation to act as an endocrine factor, explaining that apart from its key role in phosphate/calcium metabolism regulation, Klotho has also many other roles in general metabolism.

In human diseases, FGF23 as well as Klotho can be deregulated, either in genetic diseases, either in acquired diseases. Four groups can be distinguished: diseases with primary increase of FGF23 levels (e.g., hypophosphatemic rickets or tumor-induced osteomalacia), diseases with primary decrease of FGF23 levels (e.g., hyperphosphatemic tumoral calcinosis), diseases with secondary increase of FGF23 levels (e.g., chronic kidney disease, CKD), and diseases with secondary decrease of FGF23 levels (e.g., VDR deficiency leading to vitamin D-resistant rickets).

A lot of studies have focused on FGF23 and Klotho during CKD, these two biomarkers being indeed the first biomarkers to be deregulated in early CKD (increased FGF23 levels, and decreased Klotho levels). In such a disease, increased FGF23 levels have been shown to be a risk factor for cardiovascular mortality, general mortality, progression of renal disease, resistance to vitamin D analogs, and in transplant patients as a risk factor for acute rejection and graft loss.

Theoretically, the use of antiFGF23 agents in CKD patients could be of interest, but results from animal models have been disappointing, enabling the correction of hyperparathyroidism while increasing mortality. In contrast, in a next future, antiFGF23 antibodies may be of interest for genetic diseases such as hypophosphatemic rickets; therapeutic trials in this field are ongoing.

Both FGF23 and Klotho levels can be assessed in blood with immunometric assays: with the second-generation assay and its better accuracy for C-terminal FGF23, the technique is robust but unfortunately it is not available everywhere. For Klotho assessment, the assay measures the portion of α -Klotho that circulates in the systemic circulation after its release from the cell membrane. The clinical relevance of such a circulating α -Klotho remains to be fully understood, since the main biological effects of Klotho are probably mediated by the trans-membrane form of the protein.

Introduction

Fibroblast Growth Factor 23 (FGF23) is a protein synthesized mainly by osteocytes that has been described to have a key role in the 'bone-kidney' axis and the regulation of phosphate/calcium metabolism (1-3). FGF23 acts mainly as a phosphaturic factor and a suppressor of 1 α hydroxylase activity in the kidney: it inhibits the expression of type IIa and IIc sodium-phosphate cotransporters on the apical membrane of proximal tubular cells, leading to an inhibition of phosphate reabsorption (4). Moreover, it also inhibits 1 α hydroxylase activity whereas it stimulates the 24 hydroxylase activity, thus leading to decreased 1,25 dihydroxyvitamin D serum levels (5). Consequently, these two pathways account together for the hypophosphatemic effect of FGF23. FGF23 was also described as an inhibiting factor of parathyroid hormone (PTH) synthesis (6). Moreover, it is interesting to note that, in contrast to its renal effects, FGF23 can stimulate the local expression of 1 α hydroxylase in the parathyroid, suggesting that it can also indirectly down regulate PTH synthesis through an increased local production of calcitriol (7). The single-pass transmembrane Klotho protein, an anti-aging protein, seems required *in vivo* for FGF23-mediated receptor activation, at least for its renal effects (8, 9).

1 FGF23: structure and biochemical properties

FGF23 is a 251 amino-acid protein (molecular weight = 30 kDa) with a 24 amino-acid signal peptide in the N-terminal portion; its chromosomal location is 12p13 in humans. It belongs to the FGF family, in the sub-group of the 'endocrine FGFs' with FGF19 and FGF21 (10): indeed, it shares with all the FGFs a highly conserved sequence but it also has a unique C-terminal structure as well as a specific three-dimensional configuration (i.e., disulfide bound and β sheet), both accounting for its systemic action (10, 11). When it was initially described, FGF23 was thought to be cleaved by a specific metalloproteinase called PHEX (phosphate-regulating gene with homologies to endopeptidases on X chromosome) whose role has been highlighted in hypophosphatemic rickets; however, these initial findings have never been reproduced and this part of FGF23 metabolism remains to be fully understood (10, 11). However, FGF23 can be proteolytically cleaved between Arg¹⁷⁹ and Ser¹⁸⁰, in the Arg¹⁷⁶-X-X-Arg¹⁷⁹ region (4). The active form of FGF23 corresponds to the protein before cleavage, from the 25th to the 251th amino acid; in contrast, the inactive form of FGF23 is obtained after cleavage (11).

2 Animal models and FGF23

Mice knock-out for FGF23 present with a decreased longevity, in association with growth retardation, skin atrophy, decreased bone density and ectopic as well as vascular calcifications. In addition, they present with hyper-phosphatemia, hypercalcemia and increased serum levels of 1,25 dihydroxy vitamin D (12). These mice also have a trend toward an increased sensitivity to insulin and are therefore at increased risk of hypoglycemia. In these animals, a diet with low phosphate intake can improve the clinical phenotype while correcting serum phosphate levels (but without modifying serum calcium and 1,25 dihydroxy vitamin D levels). Similarly, a diet with low native vitamin D can correct serum calcium and 1,25 dihydroxy vitamin D levels (but without modifying serum phosphate levels) as well as improve life expectancy (13).

In contrast, mice over-expressing FGF23 present with a clinical phenotype of hypophosphatemic rickets, with hypophosphatemia, increased phosphaturia and hyperparathyroidism; in these animals serum calcium, 1,25 dihydroxy vitamin D circulating levels and renal function are usually normal (14). However, the renal expression of some of the regulators of phosphate reabsorption is modified, with decreased Klotho and Npt2a for example (14).

3 **Klotho: structural properties, overview on its role in physiology and mineral metabolism**

Through alternative splicing or ectodomain shedding of the transmembrane protein, the Klotho protein exists both as a secreted and a membrane protein whose extracellular domain could be shed from the cell surface by secretases and released into the circulation to act as an endocrine factor (15).

Membrane Klotho is a single-pass trans-membrane anti-aging protein (1014 amino-acids, 130 kDa, chromosomal location 13q12 in humans) that functions as a coreceptor for FGF (16). Its expression occurs mainly in the kidney and in the parathyroid. Its extra-cellular domain is wide whereas its intra-cellular part is very short.

In contrast, soluble Klotho (also known as α -Klotho) is a multifunction protein present in the biological fluids including blood, urine and cerebrospinal fluid and playing important roles in antiaging, energy metabolism, inhibition of Wnt signaling, antioxidation (through an increased endothelial nitric oxide production), resistance to lipid peroxidation, modulation of ion transport and control of PTH and vitamin D (15).

Mice over-expressing Klotho have an increased life expectancy (16). In contrast, mice lacking Klotho expression present with a decreased life span, with skin atrophy, decreased bone density, ectopic calcifications and infertility. In addition, they present with hyperphosphatemia, hypercalcemia and increased serum levels of 1,25 dihydroxy vitamin D (17). A diet with low native vitamin D can improve the whole phenotype (17). These similarities between mice knock-out for Klotho and FGF23 have led to the description of the fundamental interplay between these two players of phosphate / calcium metabolism.

Klotho is indeed an essential player for FGF23 biological activity, at least for its 'classical' effects. While FGF23 binds with a modest affinity to multiple receptors belonging to the family of FGF receptors (FGF-R, mainly type 1,3 and 4), Klotho seems required *in vivo* for FGF23-mediated receptor activation, thus stimulating the phosphorylation pathways downstream the receptor (18). The highest expression of Klotho/FGF-R complex is in the distal tubule whereas the major biologic effects of FGF23 are located in the proximal tubule (19); this discrepancy is still not explained, but other reports have also described an expression of Klotho in the proximal tubule, with a direct phosphaturic effect only by itself (20). The association between Klotho and FGF23 is stabilized *in vitro* by heparin, but its potential impact in human physiology has not been evaluated. However, FGF23 does not require Klotho in all models; indeed Klotho is not expressed in bone cells on one hand, and the effects of FGF23 are Klotho-independent in cardiomyocytes on the other hand (21).

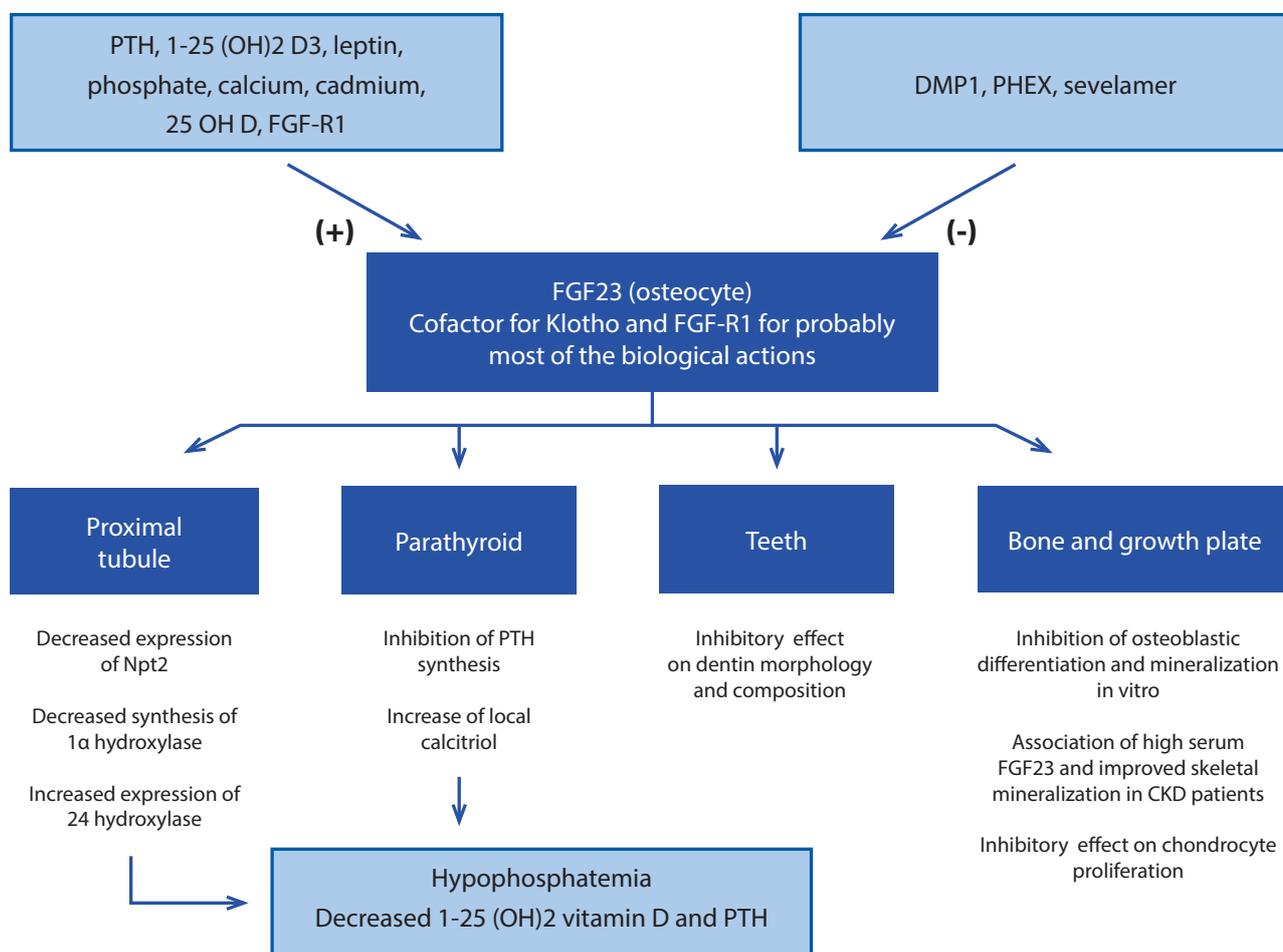
But Klotho also has its own role in the regulation of phosphate/calcium metabolism: it can function as an enzyme modifying the sugar chains of transient receptor potential vanilloid type 5 (TRPV5) in the distal tubule, preventing the calcium channel from internalization and inactivation, and thus leading to increased calcium reabsorption (22). In such a case, Klotho has a β -glucuronidase activity. Klotho can also directly regulate PTH synthesis: when intracellular calcium decreases in the parathyroid, the local expression of Klotho increases, therefore inducing an increased activity of the Na-K-ATPase channel, an increased PTH synthesis and a further correction of the hypocalcemic state (23).

Last, apart from its key role in phosphate/calcium metabolism regulation, Klotho has also many other roles in general metabolism. Klotho can regulate other ionic channels, such as the potassium channel ROMK1 (Renal Outer Medullary Potassium channel). In this later setting, its action is similar to the one observed with TRPV5: through a β -glucuronidase activity, it can stimulate urinary excretion of potassium (16). Fischer et al. have also shown that mice with Klotho deficiency have a decreased extra-cellular volume with hyperaldosteronism ; however of interest, the underlying molecular pathways remain to be proven (24). In terms of general metabolism, Klotho resists oxidative stress (through an inhibition of the IGF1 / insulin axis), modulates the Wnt pathway (16), and apoptosis through anti-apoptotic properties (25).

Although its expression is mainly located in the parathyroid and the kidney, an expression of Klotho has also been detected in other tissues, namely choroid plexus, placenta, endocrine organs such as testis, pancreas and ovary, and sinoatrial node: it is therefore likely that other new roles for Klotho will be high-lighted in the future (16). As such, α -Klotho levels were recently shown to be lower among mothers who deliver a small-for-gestational-age neonate than in those with normal pregnancies (26), and Klotho levels in the cerebrospinal fluid of patients suffering from Alzheimer's disease have been shown to be lower than in controls (27).

4 FGF23 in Human Physiology

The figure below summarizes the known roles of FGF23 in human physiology in 2013, from a 'phosphate and calcium metabolism' point of view (28-31).



(+) corresponding to a positive effect

(-) corresponding to an inhibitory effect

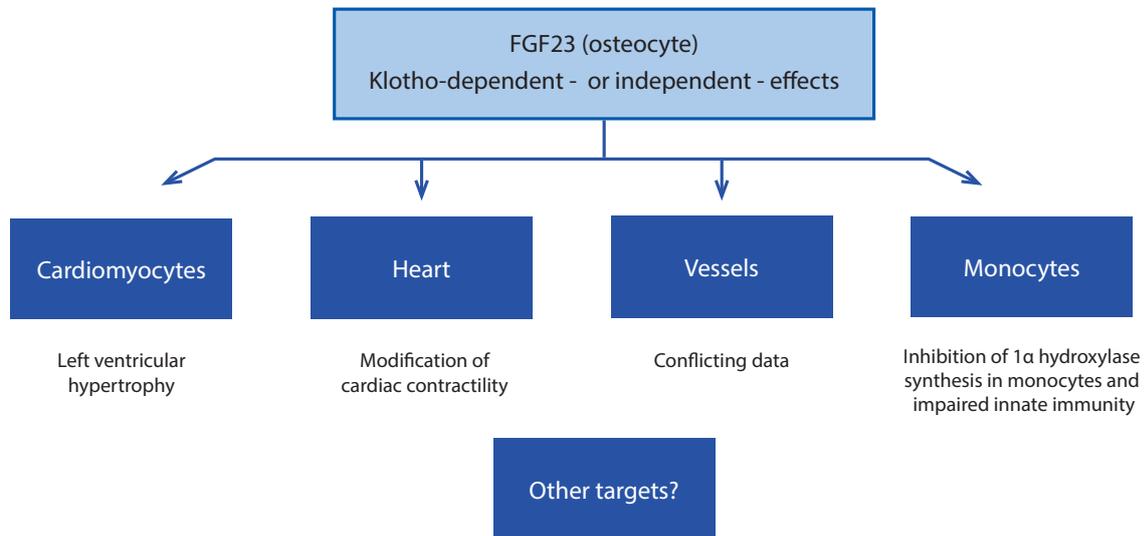
DMP1: dental matrix protein 1

PHEX: phosphate-regulating gene with homologies to endopeptidases on X chromosome

Npt 2: type II sodium-phosphate cotransporters (type a and c)

CKD: chronic kidney disease

The figure below summarizes the known roles of FGF23 in human physiology in 2013, from a more global point of view (21, 32-34).



FGF23 regulation involves both transcriptional and post-translational mechanisms, together with systemic and local bone-derived factors. PTH, vitamin D, phosphate and calcium stimulate FGF23 synthesis whereas glycoproteins synthesized by osteocytes can activate or inhibit FGF23 secretion (e.g., Matrix Extracellular Phosphoglycoprotein MEPE and Dentin Matrix Protein 1 DMP1, respectively) (35). These two proteins are strongly involved in bone mineralization, but MEPE has also been described as a phosphaturic factor by itself in the renal tubule (36). Interestingly, a recent paper also demonstrated that FGFR1 was able to regulate the FGF23 expression, an antibody-mediated activation of FGFR1 being indeed able to induce FGF23 mRNA synthesis and further hypophosphatemia in murine models; siRNA-mediated FGFR1 knockdown induced opposite effects (37).

The role of FGF23 on bone needs further evaluation but studies have demonstrated that FGF23 overexpression *in vitro* can suppress not only osteoblast differentiation but also matrix mineralization, independently of its systemic effect on phosphate metabolism (38), whereas Klotho is not expressed in bone. In contrast, other authors have reported a positive effect on FGF23 on bone: for example, Wesseling-Perry and al. have reported an association between high levels of circulating FGF23 and improved indices of skeletal mineralization (i.e., decreased osteoid thickness and shorter osteoid maturation time) in pediatric patients undergoing peritoneal dialysis (39). In medical conditions such as hypophosphatemic rickets or tumor-induced osteomalacia (TIO), the exact role of FGF23 on bone has not been yet established. A Japanese report has highlighted that patients with *osteogenesis imperfecta* receiving pamidronate infusions present an acute decrease of FGF23 circulating levels, but, again, the underlying mechanisms remain to be demonstrated (40).

5 FGF23 and Klotho in genetic human diseases

The initial description of FGF23 was done in the early 2000's, with studies focusing on hypophosphatemic rickets (41). However, it has rapidly been understood that the spectrum of FGF23 deregulation was wider, with the description of its role in TIO, and then in patients with chronic kidney disease (CKD).

Mutations of FGF23 and most of its regulators have been reported to account for either hypophosphatemic either hyperphosphatemic diseases. The table below summarizes our current knowledge of genetic diseases associated directly or indirectly (through their regulators) to FGF23 and Klotho (5, 41-45).

	DISEASE	INVOLVED GENES
Hypophosphatemia	Hypophosphatemic rickets	Activating mutation of FGF23 *** Inactivating mutation of PHEX *** Inactivating mutation of DMP1 *** Inactivating mutation of ENPP1 *** Inactivating mutation of Npt2c ** Activating translocation of Klotho * / **
	With renal lithiasis and/or osteopenia and/or hypercalciuria	Inactivating mutation of Npt2a ** Inactivating mutation of Npt2c ** Inactivating mutation of NHERF1
	Mac Cune Albright / fibrous dysplasia of bone	Overexpression of FGF23, GNAS ***
	Tumor induced osteomalacia	Overexpression of FGF23, MEPE, FGF7 and/or FRP4 ***
	Epidermal naevus syndrome	FGFR3 ***
Hyperphosphatemia	Familial tumoral calcinosis	Inactivating mutation of FGF23 ** Inactivating mutation of Klotho *** Inactivating mutation of GALNT3 **

FGF fibroblast growth factor
PHEX phosphate-regulating gene with homologies to endopeptidases on the X chromosome
DMP1 Dentin matrix protein 1
ENPP1 ecto-nucleotide pyrophosphatase / phosphodiesterase 1
Npt2a type IIa sodium-phosphate cotransporter (*SLC34A1*)
Npt2c type IIc sodium-phosphate cotransporter (*SLC34A3*)
MEPE matrix extracellular phosphoglycoprotein
FRP4 frizzled related protein 4
FGFR3 fibroblast growth factor receptor 3
GALNT3 UDP-N-acetyl- α -D-galactosamine:polypeptide N-acetylgalactosaminyltransferase 3

* with associated hyperparathyroidism
 ** disease associated with a low FGF23 serum level
 *** disease associated with a high FGF23 serum level

Briefly, hypophosphatemic rickets correspond to a heterogeneous genetic disease, affecting 1/20 000 children, resulting from mutations in FGF23, Klotho or their regulators to induce hypophosphatemia, rickets, dental abnormalities and bone deformations. Patients present with hypophosphatemia, decreased tubular phosphate reabsorption, normal 25OH vitamin D and PTH, increased alkaline phosphatase. The clinical management combines high oral phosphate intake and active vitamin D therapy, allowing a better growth and correcting the bone deformations, but explaining the two main complications often observed in this disease (i.e., secondary hyperparathyroidism secondary to phosphate intake, and nephrocalcinosis secondary to active vitamin D therapy) (46).

While hypophosphatemic rickets are usually diagnosed during early childhood, an acquired disease with the same phenotype can also affect adults (more exceptionally children or teenagers), i.e., TIO (47). These TIO correspond to mesenchymal tumors, usually benign and located in the appendicular skeleton ; they result from an acquired hypersecretion of phosphatonins (mostly FGF23, sometimes SFRP4, MEPE or FGF7) (48). Patients present with osteomalacia, bone pains, fractures and muscular weakness; a normal phosphate level years or months before the onset of the clinical picture is a strong rationale for an acquired cause of FGF23 hypersecretion. In such a case, the search for the tumor should be performed with accurate tools (tomography, MRI, Tc^{99m} bone scintigraphy, octreotide scintigraphy, F18-fluorodeoxyglucose scan), since these tumors are small and difficult to diagnose in one hand, and since hypophosphatemia will be corrected within days after surgery on the other hand (48).

In contrast, familial tumoral calcinosis is a rare autosomal recessive disorder due to inactivating mutations of FGF23, Klotho, GALTN3 or SAMD9 (49, 50). Patients present before the age of 20 with soft tissues calcifications around joints, in association with vascular calcifications and sometimes hyperostosis. Biologically, hyperphosphatemia, normal circulating calcium, increased 1-25 OH₂ vitamin D and decreased PTH levels are observed. Management is challenging, aiming at decreasing intestinal absorption of phosphorus (with sevelamer or aluminum-derived phosphate binders for example) and/or increasing urine phosphate (acetazolamide or probenecide).

6 FGF23 and Klotho deregulation during chronic kidney disease

FGF23 metabolism is strongly modified by CKD, and most studies have focused on this deregulation: FGF23 increases as GFR decreases, from early stages of CKD, even before serum phosphate and PTH have become abnormal (51, 52).

In healthy volunteers, oral phosphate loading stimulates FGF23 synthesis while it is the contrary for dietary phosphate restriction (53). FGF23 levels increase progressively as GFR decreases, some papers reporting significant increases already by stage 2 and 3, i.e., well before the onset of a critical reduction in the nephron number (54, 55). Although many studies have clearly shown the deregulation of FGF23 metabolism in the advanced stages of CKD, the mechanisms by which FGF23 serum levels increase during the early stages of CKD, even before serum phosphate or PTH increase or 1-25 vitamin D decreases, remain mysterious (56); current hypotheses propose increased FGF23 as the earliest alteration in mineral metabolism in CKD, resulting from an increased bone production rather than a decreased renal clearance (52, 54, 55). This increase could be explained by different factors, such as:

1. a decreased clearance of FGF23,
2. a compensatory mechanism in an attempt to excrete the excess serum phosphate and keep serum phosphate within the normal range,
3. a response to the treatment with active vitamin D analogs,
4. a compensatory mechanism to the loss of the kidney-secreted Klotho protein, but it is not clear whether the biological effects of FGF23 are increased or decreased in this situation, or
5. an increased production of FGF23 in bone cells (5, 42). Pereira et al. have well demonstrated in 32 children and young adults with CKD stage 2-5 that the expression of FGF23 in bone was increased at all stages of CKD, with a positive association between bone expression and circulating levels of FGF23 (31), therefore giving strength to the increased bone production hypothesis. However, the trigger of this over-production remains to be determined...

During the course of CKD, serum FGF23 levels are positively correlated with serum phosphate and negatively with serum calcitriol and PTH. The role of FGF23 to explain the onset of hyperparathyroidism can be explained by different and various direct and indirect effects. First, since FGF23 has a counter-regulatory effect on vitamin D, the increased FGF23 during CKD has the potential to reduce vitamin D activity, and thus to facilitate the development of secondary hyperparathyroidism (5). Second, FGF23 can also stimulate the local expression of 1 α hydroxylase in the parathyroid, suggesting that it could also indirectly down regulate PTH synthesis through an increased local production of calcitriol (7). Last, it has been well demonstrated both in CKD rats, in dialysis patients, and in CKD patients (among them some patients with a past of renal transplant) that there was a down-regulation of the FGF23 signaling pathway in the parathyroid glands with:

1. a decreased expression of FGFR1 and Klotho in parathyroid cells, and
2. a resistance to FGF23 administration in CKD rats, i.e., the absence of decreased PTH synthesis (57-60).

All these observations can therefore explain, at least partly, the refractory secondary hyperparathyroidism observed in CKD patients.

As detailed above, there is an accumulation of FGF23 in CKD. However, it remains questionable whether this accumulation corresponds to active or inactive fragments. In a small series of 14 end-stage renal disease (ESRD) adult patients, Weber et al. initially demonstrated an accumulation of C-term FGF23 fragments, suggesting that less than one quarter of the circulating FGF23 was bioactive in patients with end-stage renal disease, but recent data strongly support the conclusion that all circulating FGF23 in children and adults undergoing peritoneal dialysis is intact and biologically active (61). While several authors have discussed a modification of the intact/C-terminal FGF23 ratio in CKD adults (62), this ratio does not seem useless in all CKD populations (63).

In parallel to this early increase of FGF23 levels, Klotho circulating levels also decrease early during CKD (64, 65). Indeed, in this cross-sectional study of 87 CKD stage 1-5 patients, Klotho linearly decreased, whereas FGF23 showed a baseline at early CKD stages and then a curvilinear increase. Crude mean Klotho level declined by 4.8 pg/mL (95% CI 3.5-6.2 pg/mL, $P < 0.0001$) as GFR declined by 1 mL/min/1.73 m² (65). These human data were further confirmed in murine models, demonstrating a decreased renal expression of Klotho in mice with CKD (66). Moreover, Klotho polymorphisms have been associated to overall prognosis in this population: one specific Klotho polymorphism could be a protective factor against CKD progression of non-diabetic ESRD in African Americans (67), while another Klotho specific variant could increase global mortality at one year (68), and another one has been found to be associated with increased uric acid levels and decreased low density lipoprotein cholesterol in hemodialysis patients (69). Interestingly, this effect was even more marked in patients not receiving active vitamin D supplementation, keeping in mind that vitamin D is a positive regulator of Klotho (70), through the activation of a VDRE directly in the Klotho promoter. The degree of methylation of the Klotho promoter has also been shown to be associated with the severity of CKD in a cohort of Chinese patients (71). Last, in the kidney, the erythropoietin receptor and its activity are downstream effectors of Klotho enabling it to function as a cytoprotective protein against oxidative injury; these promising murine data could be of interest in the context of CKD-induced anemia (72).

All this deregulation of FGF23/Klotho are now well described in the context of chronic kidney disease, but recent data have also demonstrated an early significant increase of FGF23 levels during acute kidney injury (AKI) both in humans and in rodent models (73). In addition to this early deregulation of FGF23 in AKI, Klotho also has protective renal effects in a specific AKI model (namely cisplatin-induced AKI), through a decrease of basolateral uptake of cisplatin on one hand, and through an anti-apoptotic effect on the other hand, this later being cisplatin-independent (74).

7 FGF23, Klotho and the cardiovascular system

In general populations, Parker et al. have described an increased risk of mortality and cardiovascular events in patients with stable coronary disease and greater FGF23 levels (75), while elevated serum FGF23 levels, even within the normal range, are associated with increased left ventricular mass index and increased risk for the presence of left ventricular hypertrophy in a cohort of 795 Swedish elderly subjects (76). Recent papers also demonstrated that baseline FGF23 is a predictor of the severity of coronary artery disease in general populations (77), and that FGF23 is a predictor of severe abdominal aortic calcifications (33).

CKD patients often present with a phenotype close to the one observed in Klotho or FGF23 deficient mice, i.e., hyperphosphatemia, ectopic calcifications, vascular calcifications, hypogonadism and premature death (78). In that setting, circulating FGF23 levels can also provide prognostic information in CKD patients, mainly in terms of CKD progression, therapeutic response and cardio-vascular mortality. In a prospective cohort of 177 non diabetic CKD patients, FGF23 was an independent predictor of CKD progression, with a cut-off serum level of 104 RU/mL at inclusion for the C-term assay (79). In a prospective study of 62 dialysis patients, baseline intact FGF23 levels in association with baseline PTH serum levels were found to be good predictors of refractoriness to intravenous calcitriol therapy at 24 weeks (80). Moreover, baseline intact FGF23 serum level (cut-off 7500 ng/L) was described as a potential predictor at two years of refractory hyperparathyroidism in 103 non diabetic dialysis patients with mild hyperparathyroidism at baseline (81). Gutierrez et al. demonstrated that higher quartiles of serum FGF23 were associated with an increased risk of mortality in hemodialysis adult patients (82); Jean et al. obtained similar results in a cohort of 219 hemodialysis patients, with an increased risk of mortality and vascular calcifications in patients with higher quartiles of FGF23 serum levels two years after inclusion (83).

Recently, after these epidemiological data showing a relationship between FGF23 and cardiovascular outcomes, another step in the understanding of this pathway was made, with the first description of an off-target of FGF23 in cardiomyocytes. Indeed, Faul et al showed that FGF23 could regulate cardiomyocytes' biology in a Klotho-independent manner (21). After a clinical demonstration that the left ventricular mass index was significantly increasing with higher quartiles of circulating FGF23 levels in a prospective cohort of 3 070 adults with CKD from the CRIC study and that greater levels of FGF23 levels at baseline were also associated with a higher risk of new-onset left ventricular hypertrophy (LVH) in CKD, the authors studied the direct effects of FGF23 in cellular (cardiomyocytes from rats) and animal (wild-type mice, Klotho deficient mice and rats with 5/6th nephrectomy) models. They showed that FGF23 was able to induce *in vitro* hypertrophy of cardiomyocytes, with an activation of prohypertrophic genes; this effect was dependent of the activation of the FGF-receptors (and notably FGF-R1 and 4). Of note, in contrast to renal or parathyroid cells, these neonatal cardiomyocytes did not express Klotho. Moreover, the main downstream phosphorylation pathway involved in this activation was the PLC γ -calcineurin-NFAT axis (that can be inhibited by the use of cyclosporine), whereas in renal and parathyroid cells exposed to FGF23, the activated pathways are usually rather Erk and Akt (84). After this cellular demonstration of a direct deleterious effect of FGF23 on cardiac cells, the authors delivered FGF23 in wild-type mice either through a direct myocardial delivery of FGF23 either through an intravenous infusion for 5 days, showing in both conditions the induction of LVH. In a genetic mouse model of high FGF23 circulating levels (i.e., the Klotho deficient mice), they also showed that these mice developed LVH, therefore providing another rationale for a Klotho-independent effect of FGF23 in the cardiovascular system. Last, using a rat model of CKD (5/6th nephrectomy), they showed that a systemic treatment of these animals with the FGFR inhibitor PD173074 was able to attenuate the development of LVH, despite having no effects on the severity of CKD or arterial hypertension. More recently, Touchberry et al showed that FGF23 could also induce an elevation of intracellular calcium in cardiomyocytes, in addition to an increase of ventricular muscle strip contractility; these effects could be blocked using the L-type Ca channel blocker verapamil (32).

Thus, all these data provide a mechanistic explanation and demonstrate that FGF23 effects can be Klotho-independent, thus providing a strong rationale for future clinical trials of FGF23 monoclonal antibodies.

Taken together, all these data from epidemiological and basic studies demonstrate the presence of an association between FGF23 and left cardiac hypertrophy.

In the setting of vascular calcifications (VC), the role of FGF23 is less clear. Indeed, in the recent years of the 'FGF23 story', it was thought to be a protective factor against VC (85), but many data have challenged this initial observation. Indeed, severe calcification aortic scores (CAC) have been reported to be associated with higher FGF23 levels in non-dialysis and hemodialysis patients (86, 87), but these associations did not remain systematically significant by multivariable analyses. Scialla et al have clearly demonstrated that higher quartiles of FGF23 in CKD patients were not associated with VC while higher quartiles of phosphate were significantly associated with VC (88).

Recent experimental studies support a role for FGF23 and its co-receptor Klotho in cardiovascular pathology, yet the underlying mechanisms remain largely elusive. Klotho deficiency causes and enhances VC in murine models of CKD; indeed, Hu et al demonstrated in a model of murine CKD very low circulating and renal levels of Klotho, but CKD transgenic mice overexpressing Klotho presented with a better renal function and much less VC compared with wild-type mice with CKD (89). In a cohort of 114 CKD patients, decreased serum Klotho levels have been shown to be an independent biomarker associated with arterial stiffness; in this study Klotho levels correlated directly with 1-25 vitamin D and inversely with PTH (90).

Directly in the vessels, Klotho, FGF-R1 and FGF-R3 appear to be expressed in human normal arteries; human smooth muscle cells (normally expressing Klotho) cultured with FGF23 up-regulate the downstream kinases p-ERK and p-AKT (91). Lim et al. also described endogenous Klotho expression in human arteries and human aortic smooth muscle cells; through mechanistic studies, these authors demonstrated that Klotho knockdown potentiated the development of accelerated calcification through a Runx2 and myocardin-serum response factor-dependent pathway. Moreover, vascular cells are Klotho-dependent for FGF-23 effects since FGF23 induced the activation of p-ERK and p-AKT, which were abrogated following Klotho knockdown. Last, vascular Klotho deficiency could be restored using vitamin D receptor activators. Therefore, they described a double role for local vascular Klotho: an endogenous inhibitor of vascular calcification and a cofactor required for vascular FGF-23 signaling (92). However, Lindberg et al obtained inverse results in murine arteries from wild-type mice and from a novel mouse model harboring a vascular smooth muscle cell specific deletion of Klotho. Klotho protein levels were undetectable by immunohistochemistry and Western blots in the arteries of the two murine models. Intravenous infusion of FGF23 induced a rise in renal but not arterial Egr-1 expression, a marker of Klotho-dependent FGF23 signaling. In bovine vascular smooth muscle cells (bVSMC), the same authors demonstrated that FGF23 did not modify calcification in bVSMCs. These data therefore do not support Klotho-mediated FGF23 effects in the vasculature (93). Last, other effects of Klotho in vessels have been described such as the restoration of vasodilation capacities through endothelium-derived NO production (94), and the maintenance of endothelial integrity (95).

In the heart, Klotho is not expressed, except in the sino-atrial node where it is essential for electric activity (96). Even though there is no expression of Klotho in the heart, Klotho displays cardioprotective effects that are mediated by a downregulation of TRPC6 channels (97). Last, it is also interesting to note that an activation of the renin angiotensin system will decrease the renal expression of Klotho while the use of ACEi or ARBs will upregulate tissular Klotho (98).

In conclusion, many conflicting data have been published on the potential role of FGF23 and Klotho in heart and vessels, but FGF23 appears to be a deleterious actor in the heart while Klotho appears to have beneficial effects both in the heart and in the vessels. In the future, the next steps would be to confirm that strategies lowering FGF23 levels or increasing Klotho levels can also decrease cardiovascular and overall mortality in CKD patients; however, after the recent results published in rodents leading to increased mortality rates in CKD animals receiving the monoclonal FGF23 antibody, physicians may stay cautious (99).

8 FGF23, Klotho and global health

In terms of bone status and fracture risk, data are conflicting in the literature: some authors showed, for example in a prospective cohort of 2 868 men (75±3 years, median follow-up 3.4 years), that baseline FGF23 levels were directly correlated to the overall fracture risk, with the strongest relation when FGF23 was above 56 pg/mL; this relationship remained after adjustment on all other fracture risk factors (100). However, recent series displayed conflicting data: in the cardiovascular health study for example (corresponding to a cohort of 2 008 women and 1 329 men older than 65 years of age), greater FGF23 levels were associated with greater bone densities assessed by DXA, with no association between FGF23 and the risk of fracture (101).

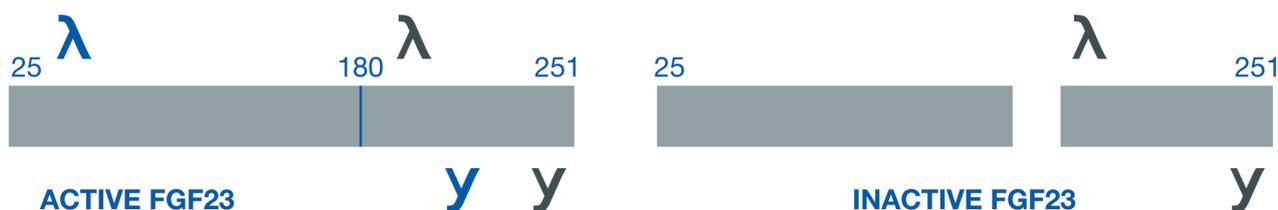
From a more metabolic point of view, other studies showed that FGF23 circulating levels were also negatively associated with high-density lipoprotein and apolipoprotein A1 and positively with triglycerides. FGF23 levels were higher in subjects with metabolic syndrome compared with those without (102). In pediatrics, FGF23 levels were also associated with BMI (103).

In humans, Klotho polymorphisms have been associated both in a positive and negative way to bone mineral density, life expectancy, cardiovascular events (e.g., ischemic stroke, carotid atherosclerosis), biomarkers of metabolic syndrome (e.g., uric acid levels, lipid and glucose metabolisms), kidney stones, and even to cognitive ability (104-110).

9 Different assays for measuring circulating FGF23 and Klotho

9.1 FGF23

Following the description of an active and an inactive form of FGF23, **different assays** have been developed for serum FGF23 measurement: the 'intact' assay that measures only active FGF23 and the 'C-terminal' assay that measures both the active and inactive FGF23. The figure below summarizes the two different types of assays for measuring FGF23 concentrations.



in blue, antibodies used to measure only the active FGF23: intact assay

in gray, antibodies used to measure both the active and the inactive FGF23: C-terminal assay

At least three studies have well demonstrated the absence of circadian intra-individual variability for FGF23 (63, 111, 112). Racial differences could account for differences in FGF23 circulating levels: for example, in a cohort of 1 099 CKD patients undergoing conservative therapies, it has been well showed after adjustment that African American patients had decreased FGF23 and 25OH vitamin D levels and increased PTH levels (113). In the same time, FGF23 levels are greater in women, in persons with tobacco exposure and diabetes (75). It remains questionable whether factors such systemic inflammation, infection or therapies can influence FGF23 serum levels by themselves; however, body weight (positive association between BMI and serum FGF23 levels), a past of solid organ transplantation as well as a corticosteroids treatment appear to influence FGF23 circulating levels (63).

Even though the manufacturers do not provide reference values, highlighting the fact that these assays should be used only for research purposes, reference values have nevertheless been proposed in healthy population: **in adults**, the reference values for C-terminal FGF23 are 55±50 RU/ml (111), and 10 to 50 pg/mL for intact FGF23 (114). There are very few **pediatric data** on FGF23. An initial study reported normal values of C-terminal FGF23 serum levels of 69±36 RU/mL in 26 children (10.9 ± 5.5 years), however without specific data confirming normal renal function

(111); since this initial report, other teams have proposed reference values according to age, gender and glomerular filtration rate with the first generation C-terminal assay and the intact one (63): the table below summarizes the reference values (results expressed as mean with 95% confidence interval) for C-terminal and intact FGF23 according to age in the 115 children with a normal renal function (i.e., glomerular filtration rate between 90 and 139 mL/min per 1.73 m²). Of note, gender did not influence the results in this pediatric population, in contrast to the Heart and Soul study (63, 75).

Age (years)	5-10	10-15	15-20
N	57	33	25
C terminal FGF23 (RU/mL)	65 (39-91)	49 (25-73)	76 (6-146)
Intact FGF23 (pg/mL)	32 (28-37)	36 (30-41)	45 (37-54)

In younger children, Brown et al. described an elevated FGF23 serum level in a child with Jansen's metaphyseal chondrodysplasia in comparison to 5 healthy children aged 10-18 months; in that study, reference FGF23 serum levels were 30±17 pg/mL with an intact assay, and 104±36 RU/mL with a C-terminal assay (115). Other papers have also studied FGF23 levels in newborns, showing a rapid decrease of the intact / C-term ratio during the first 5 days after birth (116).

In conclusion, FGF23 circulating levels on EDTA plasma can now be measured with immunometric assays that detect either the intact hormone alone or either the intact FGF23 as well as the C-terminal fragments; it seems preferable to use an intact FGF23 assay to establish the diagnosis of FGF23-dependent hypophosphatemic disorders (55).

9.2 Klotho

It is possible to measure α-Klotho circulating levels in human blood (112, 117). However, by definition, this kit measures the portion of α-Klotho that is circulating in the systemic circulation after its release from the cell membrane after shedding. The clinical relevance of such a circulating α-Klotho remains to be fully understood, since the main biological effects of Klotho are probably mediated by the trans-membrane form of the protein.

In contrast to the stability of FGF23 circulating levels among time, Klotho has a circadian rhythm, with a nadir around midnight and a maximum concentration in the early morning (112). Yamazaki et al. have reported reference serum α-Klotho values in 142 healthy volunteers (66 men, 61±19 years, serum creatinine 61±12 μmol/L): the values ranged from 239 to 1266 pg/mL (mean±SD: 562±146 pg/mL). The levels of α-Klotho were not influenced by gender or indices of skeletal metabolism, but were inversely related to serum creatinine and age. Moreover, in that study, Klotho seems to decrease with age (112, 117) and the additional analysis of 39 Asiatic children (23 boys, age 7±5 years; serum creatinine 32±11 μmol/L) showed that the concentration of the soluble form of α-Klotho was 952±282 pg/mL (i.e., significantly greater than in adults), with an intact FGF23 of 24±12 pg/mL. Analyses including both adults and children showed a positive relationship between α-Klotho and serum phosphate, whereas the association was inverse between α-Klotho and both FGF23, calcium, creatinine and age (117). Recently, Ohata et al. have also showed that the levels of alpha-Klotho were markedly higher in cord blood than in neonates at four days of life, in mothers and adult volunteers, while the fetal levels of FGF23 were lower; the levels of soluble alpha-Klotho and FGF23 were inversely correlated in cord blood (118).

In conclusion, α-Klotho circulating levels on EDTA plasma or serum can now be measured with immunometric assays but their use should be restricted for research use in 2014.

10 When should we measure FGF23 levels in clinical practice?

Before answering this question, the first question could be: should we measure FGF23 levels in clinical practice? With the second-generation assay and its better accuracy, the technique is robust but unfortunately it is not available everywhere. Two situations could theoretically benefit from FGF23 measurements: CKD-related diseases, and non-CKD diseases.

First, in non-CKD diseases, such as hypophosphatemic rickets/TIO and familial tumoral calcinosis, it could be interesting to measure FGF23 levels at the time of diagnosis; in contrast, the meaningfulness of monitoring FGF23 levels in patients with hypophosphatemic rickets receiving active vitamin D analogs and oral phosphate supplementation remains debatable since it has been shown that FGF23 levels increase with therapy, and that the alkaline phosphatase levels are a good marker of efficacy (119).

Second, in CKD, additional studies are warranted to determine whether a target of FGF23 could be proposed to adapt therapies, in addition to the calcium, phosphorus and PTH levels recommended in international guidelines depending on the different CKD stages. To date, the follow-up and drug modifications should not depend on FGF23 levels. In dialysis patients, it is probably too late to monitor FGF23 levels, it may be interesting in early CKD, but, again, simple markers of compliance are also available (such as 24-hours phosphaturia).

11 Hot topics in research

Even though many questions remain unsolved (54), FGF23 and Klotho have obviously modified our understanding of phosphate / calcium metabolism, and a therapeutic targeting of this axis will probably be the next step, at least in CKD patients. Economic and public health consequences of such strategies can be important, since the prevalence of CKD is growing worldwide, as well as the number of patients reaching ESRD.

FGF23 is a negative predictor of survival and cardio-vascular morbidity in different sub-groups of patients, but its direct toxicity has been demonstrated only in few *in vitro* models (21). However, even though it remains debatable whether FGF23 is a direct culprit or an innocent bystander (120), **strategies aiming at decreasing FGF23 levels or at restoring Klotho circulating levels could be of interest in CKD** (5, 9). In such a setting, FGF23 could also represent a biomarker to adapt all the therapies aiming at controlling CKD-MBD. Some trials have reported a decrease of FGF23 levels:

1. in early stages of CKD in adults receiving the non-calcium phosphate binder sevelamer (121),
2. in early stages of CKD in adults lowering dietary phosphate intake (122), and
3. in hemodialysis and peritoneal dialysis patients receiving the calcimimetic cinacalcet (123,124).

Moreover, a post-hoc analysis of a one-year randomized trial of phosphate binders (sevelamer vs. calcium acetate, with or without calcitriol) in 72 hemodialysis patients has showed that a more significant FGF23 decrease was obtained in patients taking sevelamer, not receiving calcitriol, and on a 2.5 mEq/l calcium dialysate (125). Other trials evaluating the effect of other non-calcium phosphate binders (i.e., lanthanum carbonate) or calcium carbonate in early CKD stages have not showed a decrease of FGF23 levels (121, 126), but a recent paper has highlighted decreased FGF23 levels in 18 patients with early CKD (stage 3) receiving lanthanum carbonate for 4 weeks (127). Moreover, in rat models of early CKD, antibodies directed against FGF23 decrease PTH levels, increase calcium and phosphate levels, and normalize 1-25 vitamin D levels (128). In healthy volunteers, native vitamin D supplementation with ergocalciferol increases FGF23 levels (129).

Will all these data in mind, the picture is nevertheless not complete, and one should not forget the results published in 2012 showing in a rat model of CKD that FGF23 neutralization using a monoclonal FGF23 antibody was able to improve the CKD-associated hyperparathyroidism by decreasing PTH levels, increasing vitamin D levels and

normalizing bone biomarkers; however, this drug increased serum phosphate, aortic calcifications and eventually the global mortality of these rats (99). This paper highlighted that we should keep in mind that the line between adaptation and maladaptation is really thin in chronic diseases (130), but also in general populations, since in healthy volunteers the administration of native vitamin D, that is commonly seen as a positive action on global health, is able to significantly increase FGF23 circulating levels (129)...

In a pediatric population, strategies aiming at decreasing FGF23 levels could also be of interest for patients with **hypophosphatemic rickets**. In a mouse model of this model (Hyp mice), the administration of specific antibodies directed against FGF23 led to an improvement of the phenotype (131, 132); human clinical trials are on-going but results are not yet available.

Recently, a link between **iron metabolism and FGF23** has also been discussed: interestingly, iron infusions (often used in CKD and dialysis patients to treat anemia in addition to erythropoietin-stimulating agents) have been showed to induce hypophosphatemia, and to increase FGF23 in patients with iron deficiency and normal renal function (133, 134). In CKD patients, one report was published, with a life-threatening hypophosphatemia following iron infusion in a woman with a past of renal transplantation (135). The clinical implications of such observations and the underlying molecular mechanisms remain to be determined, but the overall of systemic inflammation on FGF23 levels could probably be discussed. In that setting, iron deficiency may stimulate FGF23 transcription or allow FGF23 stabilization. Indeed, there is an inverse association between C-terminal FGF23 and serum iron in healthy individuals, patients with hypophosphatemic rickets and wild-type mice. Although there is also an inverse relationship between intact FGF23 and serum iron in patients with autosomal dominant hypophosphatemic rickets (ADHR, due to a mutation in the FGF23 gene leading to a resistance of FGF23 to cleavage, and thus to an accumulation of the active form of the FGF23), this later association was not found in healthy individuals, therefore leading to the hypothesis that iron could be involved in the cleavage of FGF23, but underlying molecular mechanisms beyond this phenomenon are completely unknown to date (136). Moreover, in ADHR, the human phenotype is heterogeneous, from completely unaffected patients to patients presenting with a delayed onset of the clinical hypophosphatemic phenotype; in a murine model of ADHR corresponding to a knock-in mouse with the mutation involved in FGF23 stability, it has been recently demonstrated that a low iron diet can induce the complete phenotype whereas the mice with the mutation and a normal diet were completely normal, these results thus highlighting a new example of a gene/environment interaction (137). Last, abstracts at the 2010 and 2011 American Society of Nephrology conferences as well as studies found epidemiological links between hemoglobin levels and FGF23 in CKD and in healthy persons (138).

Another point of interest would be **to determine whether FGF23 could be synthesized by other cell types in case of specific diseases**; in addition to demonstrate that baseline FGF23 levels were a predictor of mortality in patients with end-stage liver disease, it was also shown that liver cells from mice with liver lesions were able to directly express FGF23 (139). A similar mechanism could be hypothesized in patients with autosomal dominant polycystic kidney disease (ADPKD), since human data have shown that FGF23 increases in adult patients with ADPKD while circulating Klotho levels decrease, even when renal function is normal (140, 141). When comparing ADPKD patients to healthy volunteers, there was no difference for PTH, phosphorus and TmP/GFR, whereas when comparing ADPKD patients to non-diabetic CKD patients, there was no difference for PTH and TmP/GFR but a significant increase of FGF23 circulating levels and a significant decrease of phosphorus levels. With these data in mind, these authors suggest a resistance to the phosphaturic effect of FGF23 in ADPKD that could be explained through the decreased Klotho levels, but the fact that serum phosphate seemed to be lower in ADPKD patients than in healthy volunteers weakens this hypothesis. Another explanation, namely a down regulation of Klotho by increased FGF23 could also be discussed, as already shown in murine models of overexpression of FGF23 (142). Nevertheless, alternative hypotheses could be either a deregulation of FGF23 by osteocytes that could be triggered by a defect in the primary cilium complex, or an ectopic secretion of FGF23 by the tubular cells (143).

In addition to these human and animal therapeutic trials, **the underlying molecular mechanisms and cellular targets of FGF23 remain to be more accurately identified**, while Klotho is also by itself a systemic hormone with systemic and very various effects.

Other biomarkers synthesized by the ‘amazing’ osteocyte such as **sclerostin** are now widely recognized as other key factors for bone and mineral homeostasis(144), and probably also vascular calcifications ; in that setting, the recent description of a cross-regulation of vitamin D and FGF23 metabolism by sclerostin brings a new level of complexity in all this physiological mechanisms (145), but also in CKD (146, 147)...

12 Conclusion

In one decade, the description of the key role of FGF23 and Klotho in the ‘parathyroid-bone-kidney’ axis had led to a better understanding of genetic conditions associated with hypophosphatemia and phosphate calcium disorders as well as CKD-MBD. However, although a therapeutic targeting of this pathway seems likely in the future, many questions remain unresolved and conflicting; the availability of FGF23 and Klotho assays in daily practice for bench and clinical research will allow us in a next future to better understand this fascinating and global pathway.

13 References

- [1] Liu S, Quarles LD. **How fibroblast growth factor 23 works.** *J Am Soc Nephrol.* 2007 Jun;18(6):1637-47.
- [2] Yamazaki Y, Tamada T, Kasai N, Urakawa I, Aono Y, Hasegawa H, et al. **Anti-FGF23 neutralizing antibodies show the physiological role and structural features of FGF23.** *J Bone Miner Res.* 2008 Sep;23(9):1509-18.
- [3] Yoshiko Y, Wang H, Minamizaki T, Ijuin C, Yamamoto R, Suemune S, et al. **Mineralized tissue cells are a principal source of FGF23.** *Bone.* 2007 Jun;40(6):1565-73.
- [4] Fukumoto S. **Physiological regulation and disorders of phosphate metabolism--pivotal role of fibroblast growth factor 23.** *Intern Med.* 2008;47(5):337-43.
- [5] Razzaque MS. **FGF23-mediated regulation of systemic phosphate homeostasis: is Klotho an essential player?** *Am J Physiol Renal Physiol.* 2009 Mar;296(3):F470-6.
- [6] Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R, Kuro-o M, Mohammadi M, et al. **The parathyroid is a target organ for FGF23 in rats.** *J Clin Invest.* 2007 Dec;117(12):4003-8.
- [7] Krajisnik T, Bjorklund P, Marsell R, Ljunggren O, Akerstrom G, Jonsson KB, et al. **Fibroblast growth factor-23 regulates parathyroid hormone and 1alpha-hydroxylase expression in cultured bovine parathyroid cells.** *J Endocrinol.* 2007 Oct;195(1):125-31.
- [8] Kurosu H, Ogawa Y, Miyoshi M, Yamamoto M, Nandi A, Rosenblatt KP, et al. **Regulation of fibroblast growth factor-23 signaling by klotho.** *J Biol Chem.* 2006 Mar 10;281(10):6120-3.
- [9] Nakatani T, Ohnishi M, Razzaque MS. **Inactivation of klotho function induces hyperphosphatemia even in presence of high serum fibroblast growth factor 23 levels in a genetically engineered hypophosphatemic (Hyp) mouse model.** *FASEB J.* 2009 Nov;23(11):3702-11.
- [10] Fukumoto S, Yamashita T. **FGF23 is a hormone-regulating phosphate metabolism--unique biological characteristics of FGF23.** *Bone.* 2007 May;40(5):1190-5.
- [11] Yamashita T. **Structural and biochemical properties of fibroblast growth factor 23.** *Ther Apher Dial.* 2005 Aug;9(4):313-8.
- [12] Shimada T, Kakitani M, Yamazaki Y, Hasegawa H, Takeuchi Y, Fujita T, et al. **Targeted ablation of Fgf23 demonstrates an essential physiological role of FGF23 in phosphate and vitamin D metabolism.** *J Clin Invest.* 2004 Feb;113(4):561-8.
- [13] Stubbs JR, Liu S, Tang W, Zhou J, Wang Y, Yao X, et al. **Role of hyperphosphatemia and 1,25-dihydroxyvitamin D in vascular calcification and mortality in fibroblastic growth factor 23 null mice.** *J Am Soc Nephrol.* 2007 Jul;18(7):2116-24.
- [14] Marsell R, Krajisnik T, Goransson H, Ohlsson C, Ljunggren O, Larsson TE, et al. **Gene expression analysis of kidneys from transgenic mice expressing fibroblast growth factor-23.** *Nephrol Dial Transplant.* 2008 Mar;23(3):827-33.
- [15] Kovesdy CP, Quarles LD. **Fibroblast growth factor-23: what we know, what we don't know, and what we need to know.** *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association.* [Research Support, N.I.H., Extramural]. 2013 Sep;28(9):2228-36.
- [16] Kuro-o M. **Klotho.** *Pflugers Arch.* 2010 Jan;459(2):333-43.
- [17] Tsujikawa H, Kurotaki Y, Fujimori T, Fukuda K, Nabeshima Y. **Klotho, a gene related to a syndrome resembling human premature aging, functions in a negative regulatory circuit of vitamin D endocrine system.** *Mol Endocrinol.* 2003 Dec;17(12):2393-403.
- [18] Kurosu H, Kuro OM. **The Klotho gene family as a regulator of endocrine fibroblast growth factors.** *Mol Cell Endocrinol.* 2009 Feb 5;299(1):72-8.
- [18] Farrow EG, Davis SI, Summers LJ, White KE. **Initial FGF23-mediated signaling occurs in the distal convoluted tubule.** *J Am Soc Nephrol.* 2009 May;20(5):955-60.
- [20] Hu MC, Shi M, Zhang J, Pastor J, Nakatani T, Lanske B, et al. **Klotho: a novel phosphaturic substance acting as an autocrine enzyme in the renal proximal tubule.** *Faseb J.* 2010 Sep;24(9):3438-50.
- [21] Faul C, Amaral AP, Oskouei B, Hu MC, Sloan A, Isakova T, et al. **FGF23 induces left ventricular hypertrophy.** *The Journal of clinical investigation.* [Multicenter StudyResearch Support, N.I.H., ExtramuralResearch Support, Non-U.S. Gov'tResearch Support, U.S. Gov't, Non-P.H.S.]. 2011 Nov;121(11):4393-408.
- [22] Chang Q, Hoefs S, van der Kemp AW, Topala CN, Bindels RJ, Hoenderop JG. **The beta-glucuronidase klotho hydrolyzes and activates the TRPV5 channel.** *Science.* 2005 Oct 21;310(5747):490-3.
- [23] Razzaque MS. **Klotho and Na+,K+-ATPase activity: solving the calcium metabolism dilemma?** *Nephrol Dial Transplant.* 2008 Feb;23(2):459-61.
- [24] Fischer SS, Kempe DS, Leibrock CB, Rexhepaj R, Siraskar B, Boini KM, et al. **Hyperaldosteronism in Klotho-deficient mice.** *Am J Physiol Renal Physiol.* 2010 Aug 18.
- [25] Medici D, Razzaque MS, Deluca S, Rector TL, Hou B, Kang K, et al. **FGF-23-Klotho signaling stimulates proliferation and prevents vitamin D-induced apoptosis.** *J Cell Biol.* 2008 Aug 11;182(3):459-65.

- [26] Miranda J, Romero R, Korzeniewski SJ, Chaemsaitong P, Stampalija T, Yeo L, et al. **The Anti-aging Factor alpha-klotho during Human Pregnancy and its Expression in Pregnancies Complicated by Small for Gestational Age Neonates and/or Preeclampsia.** *J Matern Fetal Neona.* 2013 Jul 1.
- [27] Semba RD, Moghekar AR, Hu J, Sun K, Turner R, Ferrucci L, et al. **Klotho in the cerebrospinal fluid of adults with and without Alzheimer's disease.** *Neurosci Lett.* 2013 Nov 7.
- [28] Ben-Dov IZ, Galitzer H, Lavi-Moshayoff V, Goetz R, Kuro-o M, Mohammadi M, et al. **The parathyroid is a target organ for FGF23 in rats. The Journal of clinical investigation.** [Research Support, N.I.H., Extramural; Research Support, Non-U.S. Gov't]. 2007 Dec;117(12):4003-8.
- [29] Kawai M, Kinoshita S, Kimoto A, Hasegawa Y, Miyagawa K, Yamazaki M, et al. **FGF23 suppresses chondrocyte proliferation in the presence of soluble alpha-Klotho both in vitro and in vivo.** *The Journal of biological chemistry.* [In Vitro Research Support, Non-U.S. Gov't]. 2013 Jan 25;288(4):2414-27.
- [30] Chu EY, Fong H, Blethen FA, Tompkins KA, Foster BL, Yeh KD, et al. **Ablation of systemic phosphate-regulating gene fibroblast growth factor 23 (Fgf23) compromises the dentoalveolar complex.** *Anatomical record.* [Research Support, N.I.H., Extramural]. 2010 Jul;293(7):1214-26.
- [31] Pereira RC, Juppner H, Azucena-Serrano CE, Yadin O, Salusky IB, Wesseling-Perry K. **Patterns of FGF-23, DMP1, and MEPE expression in patients with chronic kidney disease.** *Bone.* 2009 Dec;45(6):1161-8.
- [32] Touchberry CD, Green TM, Tchikrizov V, Mannix JE, Mao TF, Carney BW, et al. **FGF23 is a novel regulator of intracellular calcium and cardiac contractility in addition to cardiac hypertrophy.** *American journal of physiology Endocrinology and metabolism.* [Research Support, N.I.H., Extramural; Research Support, Non-U.S. Gov't]. 2013 Apr 15;304(8):E863-73.
- [33] Schoppet M, Hofbauer LC, Brinselle-Schmal N, Varennes A, Goudable J, Richard M, et al. **Serum level of the phosphaturic factor FGF23 is associated with abdominal aortic calcification in men: the STRAMBO study.** *The Journal of clinical endocrinology and metabolism.* [Research Support, Non-U.S. Gov't]. 2012 Apr;97(4):E575-83.
- [34] Bacchetta J, Sea JL, Chun RF, Lisse TS, Wesseling-Perry K, Gales B, et al. **Fibroblast growth factor 23 inhibits extrarenal synthesis of 1,25-dihydroxyvitamin D in human monocytes.** *Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research.* [Research Support, N.I.H., Extramural; Research Support, Non-U.S. Gov't]. 2013 Jan;28(1):46-55.
- [35] Kawata T, Imanishi Y, Kobayashi K, Miki T, Arnold A, Inaba M, et al. **Parathyroid hormone regulates fibroblast growth factor-23 in a mouse model of primary hyperparathyroidism.** *J Am Soc Nephrol.* 2007 Oct;18(10):2683-8.
- [36] Shirley DG, Faria NJ, Unwin RJ, Dobbie H. **Direct micropuncture evidence that matrix extracellular phosphoglycoprotein inhibits proximal tubular phosphate reabsorption.** *Nephrol Dial Transplant.* 2010 Oct;25(10):3191-5.
- [37] Wu AL, Feng B, Chen MZ, Kolumam G, Zavala-Solorio J, Wyatt SK, et al. **Antibody-mediated activation of FGFR1 induces FGF23 production and hypophosphatemia.** *PLoS One.* 2013;8(2):e57322.
- [38] Wang H, Yoshiko Y, Yamamoto R, Minamizaki T, Kozai K, Tanne K, et al. **Overexpression of fibroblast growth factor 23 suppresses osteoblast differentiation and matrix mineralization in vitro.** *J Bone Miner Res.* 2008 Jun;23(6):939-48.
- [39] Wesseling-Perry K, Pereira RC, Wang H, Elashoff RM, Sahney S, Gales B, et al. **Relationship between plasma fibroblast growth factor-23 concentration and bone mineralization in children with renal failure on peritoneal dialysis.** *J Clin Endocrinol Metab.* 2009 Feb;94(2):511-7.
- [40] Kitaoka T, Namba N, Miura K, Kubota T, Ohata Y, Fujiwara M, et al. **Decrease in serum FGF23 levels after intravenous infusion of pamidronate in patients with osteogenesis imperfecta.** *J Bone Miner Metab.* 2011 Feb 23.
- [41] **Autosomal dominant hypophosphatemic rickets is associated with mutations in FGF23.** *Nat Genet.* 2000 Nov;26(3):345-8.
- [42] Prie D, Urena Torres P, Friedlander G. **Latest findings in phosphate homeostasis.** *Kidney Int.* 2009 May;75(9):882-9.
- [43] Gattineni J, Baum M. **Regulation of phosphate transport by fibroblast growth factor 23 (FGF23): implications for disorders of phosphate metabolism.** *Pediatr Nephrol.* 2010 Apr;25(4):591-601.
- [44] Lorenz-Depiereux B, Schnabel D, Tiosano D, Hausler G, Strom TM. **Loss-of-function ENPP1 mutations cause both generalized arterial calcification of infancy and autosomal-recessive hypophosphatemic rickets.** *Am J Hum Genet.* 2010 Feb 12;86(2):267-72.
- [45] Hafner C, van Oers JM, Vogt T, Landthaler M, Stoehr R, Blaszyk H, et al. **Mosaicism of activating FGFR3 mutations in human skin causes epidermal nevi.** *J Clin Invest.* 2006 Aug;116(8):2201-7.
- [46] Bacchetta J. **Rachitisme hypophosphatémique.** In: Cochat P, editor. *Progrès en Pédiatrie, Néphrologie Pédiatrique:* Doin; 2010. p. Epub.
- [47] Reyes-Mugica M, Arnsmeier SL, Backeljauw PF, Persing J, Ellis B, Carpenter TO. **Phosphaturic mesenchymal tumor-induced rickets.** *Pediatr Dev Pathol.* 2000 Jan-Feb;3(1):61-9.
- [48] Woo VL, Landesberg R, Imel EA, Singer SR, Folpe AL, Econs MJ, et al. **Phosphaturic mesenchymal tumor, mixed connective tissue variant, of the mandible: report of a case and review of the literature.** *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2009 Dec;108(6):925-32.
- [49] Ichikawa S, Imel EA, Kreiter ML, Yu X, Mackenzie DS, Sorenson AH, et al. **A homozygous missense mutation in human KLOTHO causes severe tumoral calcinosis.** *The Journal of clinical investigation.* [Case Reports; Research Support, N.I.H., Extramural; Research Support, Non-U.S. Gov't]. 2007 Sep;117(9):2684-91.
- [50] **Uptake, distribution, and esterification of retinol in liver.** *Nutr Rev.* 1986 Jun;44(6):216-7.
- [51] Danziger J. **The bone-renal axis in early chronic kidney disease: an emerging paradigm.** *Nephrol Dial Transplant.* 2008 Sep;23(9):2733-7.
- [52] Isakova T, Wahl P, Vargas GS, Gutierrez OM, Scialla J, Xie H, et al. **Fibroblast growth factor 23 is elevated before parathyroid hormone and phosphate in chronic kidney disease.** *Kidney Int.* 2011 Mar 9.
- [53] Burnett SM, Gunawardene SC, Bringhurst FR, Juppner H, Lee H, Finkelstein JS. **Regulation of C-terminal and intact FGF-23 by dietary phosphate in men and women.** *J Bone Miner Res.* 2006 Aug;21(8):1187-96.
- [54] Wolf M. **Forging forward with 10 burning questions on FGF23 in kidney disease.** *J Am Soc Nephrol.* 2010 Sep;21(9):1427-35.
- [55] Juppner H, Wolf M, Salusky IB. **FGF23: more than a regulator of renal phosphate handling?** *J Bone Miner Res.* 2010 Jun 30.
- [56] Isakova T, Wolf MS. **FGF23 or PTH: which comes first in CKD?** *Kidney Int.* 2011 Nov;78(10):947-9.
- [57] Lafage-Proust MH. **Does the downregulation of the FGF23 signaling pathway in hyperplastic parathyroid glands contribute to refractory secondary hyperparathyroidism in CKD patients?** *Kidney Int.* 2010 Mar;77(5):390-2.
- [58] Komaba H, Goto S, Fujii H, Hamada Y, Kobayashi A, Shibuya K, et al. **Depressed expression of Klotho and FGF receptor 1 in hyperplastic parathyroid glands from uremic patients.** *Kidney Int.* 2010 Feb;77(3):232-8.
- [59] Galitzer H, Ben-Dov IZ, Silver J, Naveh-Many T. **Parathyroid cell resistance to fibroblast growth factor 23 in secondary hyperparathyroidism of chronic kidney disease.** *Kidney Int.* 2010 Feb;77(3):211-8.
- [60] Krajcnsnik T, Olauson H, Mirza MA, Hellman P, Akerstrom G, Westin G, et al. **Parathyroid Klotho and FGF-receptor 1 expression decline with renal function in hyperparathyroid patients with chronic kidney disease and kidney transplant recipients.** *Kidney Int.* 2010 Nov;78(10):1024-32.
- [61] Shimada T, Urakawa I, Isakova T, Yamazaki Y, Epstein M, Wesseling-Perry K, et al. **Circulating Fibroblast Growth Factor 23 in Patients with End-Stage Renal Disease Treated by Peritoneal Dialysis Is Intact and Biologically Active.** *J Clin Endocrinol Metab.* 2009 Dec 4.
- [62] Imanishi Y, Inaba M, Nakatsuka K, Nagasue K, Okuno S, Yoshihara A, et al. **FGF-23 in patients with end-stage renal disease on hemodialysis.** *Kidney Int.* 2004 May;65(5):1943-6.

- [63] Bacchetta J, Dubourg L, Harambat J, Ranchin B, Abou-Jaoude P, Arnaud S, et al. **The influence of glomerular filtration rate and age on fibroblast growth factor 23 serum levels in pediatric chronic kidney disease.** *J Clin Endocrinol Metab.* 2010 Apr;95(4):1741-8.
- [64] Hu MC, Kuro-o M, Moe OW. **The emerging role of Klotho in clinical nephrology.** *Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association - European Renal Association.* [Research Support, N.I.H., Extramural; Research Support, Non-U.S. Gov't]. 2012 Jul;27(7):2650-7.
- [65] Pavik I, Jaeger P, Ebner L, Wagner CA, Petzold K, Spichtig D, et al. **Secreted Klotho and FGF23 in chronic kidney disease Stage 1 to 5: a sequence suggested from a cross-sectional study.** *Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association - European Renal Association.* [Research Support, Non-U.S. Gov't]. 2013 Feb;28(2):352-9.
- [66] Zhang S, Gillihan R, He N, Fields T, Liu S, Green T, et al. **Dietary phosphate restriction suppresses phosphaturia but does not prevent FGF23 elevation in a mouse model of chronic kidney disease.** *Kidney international.* 2013 May 22.
- [67] Bostrom MA, Hicks PJ, Lu L, Langefeld CD, Freedman BI, Bowden DW. **Association of polymorphisms in the Klotho gene with severity of non-diabetic ESRD in African Americans.** *Nephrol Dial Transplant.* 2010 Oct;25(10):3348-55.
- [68] Friedman DJ, Afkarian M, Tamez H, Bhan I, Isakova T, Wolf M, et al. **Klotho Variants and Chronic Hemodialysis Mortality.** *J Bone Miner Res.* 2009 May 6.
- [69] Shimoyama Y, Taki K, Mitsuda Y, Tsuruta Y, Hamajima N, Niwa T. **KLOTHO gene polymorphisms G-395A and C1818T are associated with low-density lipoprotein cholesterol and uric acid in Japanese hemodialysis patients.** *Am J Nephrol.* 2009;30(4):383-8.
- [70] Kido S, Kaneko I, Tatsumi S, Segawa H, Miyamoto K. **Vitamin D and type II sodium-dependent phosphate cotransporters.** *Contrib Nephrol.* 2013;180:86-97.
- [71] Chen J, Zhang X, Zhang H, Lin J, Zhang C, Wu Q, et al. **Elevated klotho promoter methylation is associated with severity of chronic kidney disease.** *PLoS One.* 2013;8(11):e79856.
- [72] Hu MC, Shi M, Cho HJ, Zhang J, Pavlenco A, Liu S, et al. **The erythropoietin receptor is a downstream effector of Klotho-induced cytoprotection.** *Kidney international.* 2013 May 1.
- [73] Christov M, Waikar SS, Pereira RC, Havasi A, Leaf DE, Goltzman D, et al. **Plasma FGF23 levels increase rapidly after acute kidney injury.** *Kidney international.* 2013 May 8.
- [74] Panesso MC, Shi M, Cho HJ, Paek J, Ye J, Moe OW, et al. **Klotho has dual protective effects on cisplatin-induced acute kidney injury.** *Kidney international.* 2013 Dec 4.
- [75] Parker BD, Schurgers LJ, Brandenburg VM, Christenson RH, Vermeer C, Ketteler M, et al. **The associations of fibroblast growth factor 23 and uncarboxylated matrix Gla protein with mortality in coronary artery disease: the Heart and Soul Study.** *Ann Intern Med.* 2010 May 18;152(10):640-8.
- [76] Mirza MA, Larsson A, Lind L, Larsson TE. **Circulating fibroblast growth factor-23 is associated with vascular dysfunction in the community.** *Atherosclerosis.* 2009 Jan 9.
- [77] Chen YC, Lin FY, Lin RH, Chuang CL, Chang CC, Tsai CS. **Relation Between Fetuin-A Levels and Fibroblast Growth Factor 23 With the Severity of Coronary Artery Disease Measured by SYNTAX Scores.** *The American journal of cardiology.* 2013 Jun 21.
- [78] Razaque MS, Lanske B. **Hypervitaminosis D and premature aging: lessons learned from Fgf23 and Klotho mutant mice.** *Trends Mol Med.* 2006 Jul;12(7):298-305.
- [79] Fliser D, Kollerits B, Neyer U, Ankerst DP, Lhotta K, Lingenhel A, et al. **Fibroblast growth factor 23 (FGF23) predicts progression of chronic kidney disease: the Mild to Moderate Kidney Disease (MMKD) Study.** *J Am Soc Nephrol.* 2007 Sep;18(9):2600-8.
- [80] Kazama JJ, Sato F, Omori K, Hama H, Yamamoto S, Maruyama H, et al. **Pretreatment serum FGF-23 levels predict the efficacy of calcitriol therapy in dialysis patients.** *Kidney Int.* 2005 Mar;67(3):1120-5.
- [81] Nakanishi S, Kazama JJ, Nii-Kono T, Omori K, Yamashita T, Fukumoto S, et al. **Serum fibroblast growth factor-23 levels predict the future refractory hyperparathyroidism in dialysis patients.** *Kidney Int.* 2005 Mar;67(3):1171-8.
- [82] Gutierrez OM, Mannstadt M, Isakova T, Rauh-Hain JA, Tamez H, Shah A, et al. **Fibroblast growth factor 23 and mortality among patients undergoing hemodialysis.** *N Engl J Med.* 2008 Aug 7;359(6):584-92.
- [83] Jean G, Terrat JC, Vanel T, Hurot JM, Lorriaux C, Mayor B, et al. **High levels of serum fibroblast growth factors (FGF)-23 are associated with increased mortality in long haemodialysis patients.** *Nephrol Dial Transplant.* 2009 Apr 25.
- [84] Andrukhova O, Zeitz U, Goetz R, Mohammadi M, Lanske B, Erben RG. **FGF23 acts directly on renal proximal tubules to induce phosphaturia through activation of the ERK1/2-SGK1 signaling pathway.** *Bone.* 2012 May 27.
- [85] Inaba M, Okuno S, Imanishi Y, Yamada S, Shioi A, Yamakawa T, et al. **Role of fibroblast growth factor-23 in peripheral vascular calcification in non-diabetic and diabetic hemodialysis patients.** *Osteoporosis international: a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA.* 2006 Oct;17(10):1506-13.
- [86] Morena M, Jausset I, Halkovich A, Dupuy AM, Bargnoux AS, Chenine L, et al. **Bone biomarkers help grading severity of coronary calcifications in non dialysis chronic kidney disease patients.** *PLoS One.* [Research Support, Non-U.S. Gov't]. 2012;7(5):e36175.
- [87] Desjardins L, Liabeuf S, Renard C, Lenglet A, Lemke HD, Choukroun G, et al. **FGF23 is independently associated with vascular calcification but not bone mineral density in patients at various CKD stages.** *Osteoporosis international: a journal established as result of cooperation between the European Foundation for Osteoporosis and the National Osteoporosis Foundation of the USA.* 2012 Jul;23(7):2017-25.
- [88] Scialla JJ, Lau WL, Reilly MP, Isakova T, Yang HY, Crouthamel MH, et al. **Fibroblast growth factor 23 is not associated with and does not induce arterial calcification.** *Kidney international.* [Research Support, N.I.H., Extramural]. 2013 Jun;83(6):1159-68.
- [89] Hu MC, Shi M, Zhang J, Quinones H, Griffith C, Kuro-o M, et al. **Klotho deficiency causes vascular calcification in chronic kidney disease.** *Journal of the American Society of Nephrology: JASN.* [Research Support, N.I.H., Extramural; Research Support, Non-U.S. Gov't]. 2011 Jan;22(1):124-36.
- [90] Kitagawa M, Sugiyama H, Morinaga H, Inoue T, Takiue K, Ogawa A, et al. **A decreased level of serum soluble Klotho is an independent biomarker associated with arterial stiffness in patients with chronic kidney disease.** *PLoS One.* 2013;8(2):e56695.
- [91] Lim K LT, Zehnder D, Hsiao LL. **Development of Klotho-FGFR1/3 dependent resistance to FGF23 in human aortic smooth muscle cells exposed to calcifying stress.** *American Society of Nephrology, Denver, PO126.* 2010.
- [92] Lim K, Lu TS, Molostvov G, Lee C, Lam FT, Zehnder D, et al. **Vascular Klotho deficiency potentiates the development of human artery calcification and mediates resistance to fibroblast growth factor 23.** *Circulation.* [Research Support, Non-U.S. Gov't]. 2012 May 8;125(18):2243-55.
- [93] Lindberg K, Olauson H, Amin R, Ponnusamy A, Goetz R, Taylor RF, et al. **Arterial klotho expression and FGF23 effects on vascular calcification and function.** *PLoS One.* [Research Support, Non-U.S. Gov't]. 2013;8(4):e60658.
- [94] Saito Y, Yamagishi T, Nakamura T, Ohyama Y, Aizawa H, Suga T, et al. **Klotho protein protects against endothelial dysfunction. Biochemical and biophysical research communications.** [Research Support, Non-U.S. Gov't]. 1998 Jul 20;248(2):324-9.
- [95] Kusaba T, Okigaki M, Matui A, Murakami M, Ishikawa K, Kimura T, et al. **Klotho is associated with VEGF receptor-2 and the transient receptor potential canonical-1 Ca2+ channel to maintain endothelial integrity.** *Proceedings of the National Academy of Sciences of the United States of America.* [Research Support, Non-U.S. Gov't]. 2010 Nov 9;107(45):19308-13.

- [95] Takeshita K, Fujimori T, Kurotaki Y, Honjo H, Tsujikawa H, Yasui K, et al. **Sinoatrial node dysfunction and early unexpected death of mice with a defect of klotho gene expression.** *Circulation*. 2004 Apr 13;109(14):1776-82.
- [95] Xie J, Cha SK, An SW, Kuro OM, Birnbaumer L, Huang CL. **Cardioprotection by Klotho through downregulation of TRPC6 channels in the mouse heart.** *Nat Commun*. [Research Support, N.I.H., Extramural; Research Support, N.I.H., Intramural; Research Support, Non-U.S. Gov't]. 2012;3:1238.
- [95] de Borst MH, Vervloet MG, ter Wee PM, Navis G. **Cross talk between the renin-angiotensin-aldosterone system and vitamin D-FGF-23-klotho in chronic kidney disease.** *Journal of the American Society of Nephrology : JASN*. [Research Support, Non-U.S. Gov't]. 2011 Sep;22(9):1603-9.
- [99] Shalhoub V, Shatzken EM, Ward SC, Davis J, Stevens J, Bi V, et al. **FGF23 neutralization improves chronic kidney disease-associated hyperparathyroidism yet increases mortality.** *The Journal of clinical investigation*. [Research Support, Non-U.S. Gov't]. 2012 Jul 2;122(7):2543-53.
- [100] Mirza MA, Karlsson MK, Mellstrom D, Orwoll E, Ohlsson C, Ljunggren O, et al. **Serum fibroblast growth factor-23 (FGF23) and fracture risk in elderly men.** *J Bone Miner Res*. 2010 Oct 6.
- [101] Jovanovich A, Buzkova P, Chonchol M, Robbins J, Fink HA, de Boer IH, et al. **Fibroblast Growth Factor 23, Bone Mineral Density, and Risk of Hip Fracture among Older Adults: The Cardiovascular Health Study.** *The Journal of clinical endocrinology and metabolism*. 2013 Jun 14.
- [102] Mirza MA, Alsio J, Hammarstedt A, Erben RG, Michaelsson K, Tivesten A, et al. **Circulating fibroblast growth factor-23 is associated with fat mass and dyslipidemia in two independent cohorts of elderly individuals.** *Arterioscler Thromb Vasc Biol*. 2011 Jan;31(1):219-27.
- [103] Bacchetta J, Dubourg L, Harambat J, Ranchin B, Abou-Jaoude P, Arnaud S, et al. **The Influence of Glomerular Filtration Rate and Age on Fibroblast Growth Factor 23 Serum Levels in Pediatric Chronic Kidney Disease.** *J Clin Endocrinol Metab*. 2010 Feb 15.
- [104] Majumdar V, Nagaraja D, Christopher R. **Association of the functional KL-VS variant of Klotho gene with early-onset ischemic stroke.** *Biochem Biophys Res Commun*. 2010 Dec 17;403(3-4):412-6.
- [105] Oguro R, Kamide K, Kokubo Y, Shimaoka I, Congrains A, Horio T, et al. **Association of carotid atherosclerosis with genetic polymorphisms of the klotho gene in patients with hypertension.** *Geriatr Gerontol Int*. 2011 Oct;10(4):311-8.
- [106] Shimoyama Y, Nishio K, Hamajima N, Niwa T. **KLOTHO gene polymorphisms G-395A and C1818T are associated with lipid and glucose metabolism, bone mineral density and systolic blood pressure in Japanese healthy subjects.** *Clin Chim Acta*. 2009 Aug;406(1-2):134-8.
- [107] Zarrabeitia MT, Hernandez JL, Valero C, Zarrabeitia AL, Ortiz F, Gonzalez-Macias J, et al. **Klotho gene polymorphism and male bone mass.** *Calcif Tissue Int*. 2007 Jan;80(1):10-4.
- [108] Deary IJ, Harris SE, Fox HC, Hayward C, Wright AF, Starr JM, et al. **KLOTHO genotype and cognitive ability in childhood and old age in the same individuals.** *Neurosci Lett*. 2005 Apr 11;378(1):22-7.
- [109] Bonafe M, Olivieri F. **Genetic polymorphism in long-lived people: cues for the presence of an insulin/IGF-pathway-dependent network affecting human longevity.** *Mol Cell Endocrinol*. 2009 Feb 5;299(1):118-23.
- [110] Telci D, Dogan AU, Ozbek E, Polat EC, Simsek A, Cakir SS, et al. **KLOTHO gene polymorphism of G395A is associated with kidney stones.** *American journal of nephrology*. 2011;33(4):337-43.
- [111] Jonsson KB, Zahradnik R, Larsson T, White KE, Sugimoto T, Imanishi Y, et al. **Fibroblast growth factor 23 in oncogenic osteomalacia and X-linked hypophosphatemia.** *N Engl J Med*. 2003 Apr 24;348(17):1656-63.
- [112] Carpenter TO, Insogna KL, Zhang JH, Ellis B, Nieman S, Simpson C, et al. **Circulating Levels of Soluble Klotho and FGF23 in X-Linked Hypophosphatemia: Circadian Variance, Effects of Treatment and Relationship to Parathyroid Status.** *J Clin Endocrinol Metab*. 2010 Aug 4.
- [113] Jovanovich AJ CM, Cheung AK, Kaufman JS, Greene TH, Roberts WL, Smits GJ, Kendrick JB. **Racial differences in vitamin D, parathyroid hormone and FGF23 in patients with severe chronic kidney disease.** *American Society of Nephrology, Denver, PO2153*. 2010.
- [114] Yamazaki Y, Okazaki R, Shibata M, Hasegawa Y, Satoh K, Tajima T, et al. **Increased circulatory level of biologically active full-length FGF-23 in patients with hypophosphatemic rickets/osteomalacia.** *J Clin Endocrinol Metab*. 2002 Nov;87(11):4957-60.
- [115] Brown WW, Juppner H, Langman CB, Price H, Farrow EG, White KE, et al. **Hypophosphatemia with elevations in serum fibroblast growth factor 23 in a child with Jansen's metaphyseal chondrodysplasia.** *J Clin Endocrinol Metab*. 2009 Jan;94(1):17-20.
- [116] Takaiwa M, Aya K, Miyai T, Hasegawa K, Yokoyama M, Kondo Y, et al. **Fibroblast growth factor 23 concentrations in healthy term infants during the early postpartum period.** *Bone*. Aug;47(2):256-62.
- [117] Yamazaki Y, Imura A, Urakawa I, Shimada T, Murakami J, Aono Y, et al. **Establishment of sandwich ELISA for soluble alpha-Klotho measurement: Age-dependent change of soluble alpha-Klotho levels in healthy subjects.** *Biochem Biophys Res Commun*. 2010 Jul 30;398(3):513-8.
- [118] Ohata Y, Arahori H, Namba N, Kitaoka T, Hirai H, Wada K, et al. **Circulating Levels of Soluble (alpha)-Klotho Are Markedly Elevated in Human Umbilical Cord Blood.** *J Clin Endocrinol Metab*. 2011 Mar 16.
- [119] Imel EA, DiMeglio LA, Hui SL, Carpenter TO, Econs MJ. **Treatment of X-linked hypophosphatemia with calcitriol and phosphate increases circulating fibroblast growth factor 23 concentrations.** *The Journal of clinical endocrinology and metabolism*. [Clinical Trial; Research Support, N.I.H., Extramural]. 2010 Apr;95(4):1846-50.
- [120] Razzaque MS. **Does FGF23 toxicity influence the outcome of chronic kidney disease?** *Nephrol Dial Transplant*. 2009 Jan;24(1):4-7.
- [121] Oliveira RB, Cancela AL, Gracioli FG, Dos Reis LM, Draibe SA, Cuppari L, et al. **Early control of PTH and FGF23 in normophosphatemic CKD patients: a new target in CKD-MBD therapy?** *Clin J Am Soc Nephrol*. 2010 Feb;5(2):286-91.
- [122] Sigrist MK, Beaulieu MC, A L. **Lowering dietary phosphate intake reduces FGF23 in CKD stages 3 and 4.** *American Society of Nephrology, Denver, PO165*. 2010.
- [123] Koizumi M, Komaba H, Tanaka H, Nakanishi S, Fujimori A, M F. **Cinacalcet for secondary hyperparathyroidism lowers FGF23 levels in patients receiving hemodialysis.** *American Society of Nephrology, Denver, PO165*. 2010.
- [124] Kim HJ, Kim H, Shin N, Na KY, Kim YL, Kim D, et al. **Cinacalcet lowering of serum fibroblast growth factor-23 concentration may be independent from serum Ca, P, PTH and dose of active vitamin D in peritoneal dialysis patients: a randomized controlled study.** *BMC Nephrol*. 2013;14:112.
- [125] Cancela AL, Oliveira RB, Gracioli FG, dos Reis LM, Barreto F, Barreto DV, et al. **Fibroblast growth factor 23 in hemodialysis patients: effects of phosphate binder, calcitriol and calcium concentration in the dialysate.** *Nephron Clin Pract*. 2010;117(1):c74-82.
- [126] Isakova T, Gutierrez OM, Smith K, Epstein M, Keating LK, Juppner H, et al. **Pilot study of dietary phosphorus restriction and phosphorus binders to target fibroblast growth factor 23 in patients with chronic kidney disease.** *Nephrol Dial Transplant*. 2011 Feb;26(2):584-91.
- [127] Gonzalez-Parra E, Gonzalez-Casaus ML, Galan A, Martinez-Calero A, Navas V, Rodriguez M, et al. **Lanthanum carbonate reduces FGF23 in chronic kidney disease stage 3 patients.** *Nephrol Dial Transplant*. 2011 Mar 24.
- [128] Hasegawa H, Nagano N, Urakawa I, Yamazaki Y, Iijima K, Fujita T, et al. **Direct evidence for a causative role of FGF23 in the abnormal renal phosphate handling and vitamin D metabolism in rats with early-stage chronic kidney disease.** *Kidney Int*. 2010 Nov;78(10):975-80.

- [129] Burnett-Bowie SA, Leder BZ, Henao MP, Baldwin CM, Hayden DL, Finkelstein JS. **Randomized trial assessing the effects of ergocalciferol administration on circulating FGF23.** *Clinical journal of the American Society of Nephrology : CJASN.* [Randomized Controlled Trial Research Support, N.I.H., Extramural Research Support, Non-U.S. Gov't]. 2012 Apr;7(4):624-31.
- [130] Ketteler M, Biggar PH, Liangos O. **FGF23 antagonism: the thin line between adaptation and maladaptation in chronic kidney disease.** *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association.* 2013 Apr;28(4):821-5.
- [131] Aono Y, Yamazaki Y, Yasutake J, Kawata T, Hasegawa H, Urakawa I, et al. **Therapeutic effects of anti-FGF23 antibodies in hypophosphatemic rickets/osteomalacia.** *J Bone Miner Res.* 2009 Nov;24(11):1879-88.
- [132] Aono Y, Hasegawa H, Yamazaki Y, Shimada T, Fujita T, Yamashita T, et al. **Anti-FGF23 neutralizing antibodies ameliorate muscle weakness and decreased spontaneous movement of Hyp mice.** *J Bone Miner Res.* 2010 Oct 11.
- [133] Schouten BJ, Hunt PJ, Livesey JH, Frampton CM, Soule SG. **FGF23 elevation and hypophosphatemia after intravenous iron polymaltose: a prospective study.** *J Clin Endocrinol Metab.* 2009 Jul;94(7):2332-7.
- [134] Shimizu Y, Tada Y, Yamauchi M, Okamoto T, Suzuki H, Ito N, et al. **Hypophosphatemia induced by intravenous administration of saccharated ferric oxide: another form of FGF23-related hypophosphatemia.** *Bone.* 2009 Oct;45(4):814-6.
- [135] Mani LY, Nseir G, Venetz JP, Pascual M. **Severe hypophosphatemia after intravenous administration of iron carboxymaltose in a stable renal transplant recipient.** *Transplantation.* 2010 Oct 15;90(7):804-5.
- [136] Imel EA, Peacock M, Gray AK, Padgett LR, Hui SL, Econs MJ. **Iron modifies plasma FGF23 differently in autosomal dominant hypophosphatemic rickets and healthy humans.** *The Journal of clinical endocrinology and metabolism.* [Research Support, N.I.H., Extramural]. 2011 Nov;96(11):3541-9.
- [137] Farrow EG, Yu X, Summers LJ, Davis SI, Fleet JC, Allen MR, et al. **Iron deficiency drives an autosomal dominant hypophosphatemic rickets (ADHR) phenotype in fibroblast growth factor-23 (Fgf23) knock-in mice.** *Proceedings of the National Academy of Sciences of the United States of America.* [Research Support, N.I.H., Extramural Research Support, Non-U.S. Gov't]. 2011 Nov 15;108(46):E1146-55.
- [138] Braithwaite V, Jarjou LM, Goldberg GR, Prentice A. **Iron status and fibroblast growth factor-23 in Gambian children.** *Bone [Research Support, Non-U.S. Gov't].* 2012 Jun;50(6):1351-6.
- [139] Prie D, Forand A, Francoz C, Elie C, Cohen I, Courbebaisse M, et al. **Plasma fibroblast growth factor 23 concentration is increased and predicts mortality in patients on the liver-transplant waiting list.** *PLoS One.* 2013;8(6):e66182.
- [140] Pavik I, Jaeger P, Kistler AD, Poster D, Krauer F, Cavelti-Weder C, et al. **Patients with autosomal dominant polycystic kidney disease have elevated fibroblast growth factor 23 levels and a renal leak of phosphate.** *Kidney international.* [Research Support, Non-U.S. Gov't]. 2011 Jan;79(2):234-40.
- [141] Pavik I, Jaeger P, Ebner L, Poster D, Krauer F, Kistler AD, et al. **Soluble klotho and autosomal dominant polycystic kidney disease.** *Clinical journal of the American Society of Nephrology: CJASN.* [Research Support, Non-U.S. Gov't]. 2012 Feb;7(2):248-57.
- [142] Marsell R, Krajisnik T, Goransson H, Ohlsson C, Ljunggren O, Larsson TE, et al. **Gene expression analysis of kidneys from transgenic mice expressing fibroblast growth factor-23.** *Nephrology, dialysis, transplantation : official publication of the European Dia2008 Mar;23(3):827-33.*
- [143] Mekahli D, Bacchetta J. **From bone abnormalities to mineral metabolism dysregulation in autosomal dominant polycystic kidney disease.** *Pediatric nephrology.* 2013 Jan 24.
- [144] Bonewald LF. **The amazing osteocyte.** *Journal of bone and mineral research : the official journal of the American Society for Bone and Mineral Research.* [Research Support, N.I.H., Extramural Review]. 2011 Feb;26(2):229-38.
- [145] Ryan ZC, Ketha H, McNulty MS, McGee-Lawrence M, Craig TA, Grande JP, et al. **Sclerostin alters serum vitamin D metabolite and fibroblast growth factor 23 concentrations and the urinary excretion of calcium.** *Proceedings of the National Academy of Sciences of the United States of America.* 2013 Apr 9;110(15):6199-204.
- [146] Viaene L, Behets GJ, Claes K, Meijers B, Blocki F, Brandenburg V, et al. **Sclerostin: another bone-related protein related to all-cause mortality in haemodialysis?** *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association - European Renal Association.* 2013 Apr 19.
- [147] Pelletier S, Dubourg L, Carlier MC, Hadj-Aissa A, Fouque D. **The relation between renal function and serum sclerostin in adult patients with CKD.** *Clinical journal of the American Society of Nephrology : CJASN.* 2013 May;8(5):819-23.

FGF-23 Intact, Human 2nd Generation

FIBROBLAST GROWTH FACTOR 23

Intact

Cat. No. 60 - 6600

Tests 96

Method ELISA

Range 20 - 660 pg/ml (can be extended to 2200 pg/ml)

Sensitivity 1.5 pg/ml

Incubation time 3.5 hours

Sample volume 50 µl

Sample type EDTA & Heparin plasma, cell culture

Sample preparation It is recommended to collect the sample in the morning after a 12-hour fasting period. Intact FGF-23 is very instable. Therefore, collection and testing or storage should take place promptly. Store samples at -20 °C or below. Avoid repeated freezing and thawing of specimens.

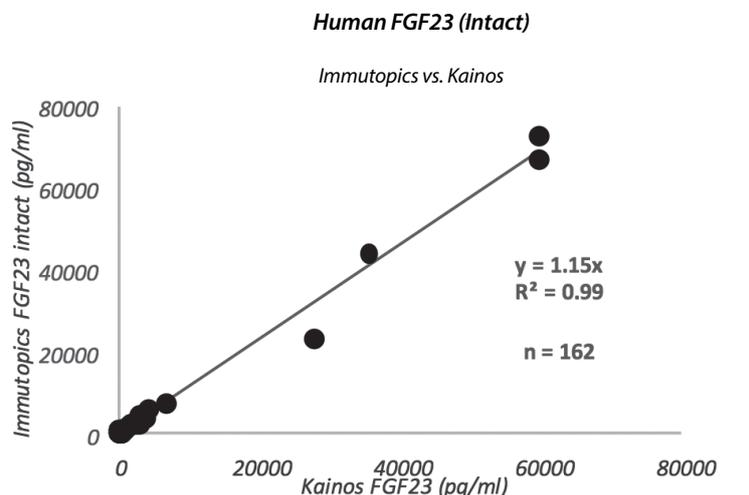
Reference values Mean: 40.7 pg/ml, SD 16.1 pg/ml

Range: 8.6 - 72.8 pg/ml +/- 2 SD

Species Human

Specificity Antibodies bind to NH2-terminal and C-terminal epitopes; measures only the intact form of the molecule.

Correlation



FGF-23 (C-Term) 2nd Generation

FIBROBLAST GROWTH FACTOR 23

C-terminal

Cat. No. 60 - 6100

Tests 96

Method

Range 1.5 - 1500 RU/ml

Sensitivity 1.5 RU/ml

Incubation time 3.5 hours

Sample volume 100 µl

Sample type Heparin- and EDTA Plasma, cell culture

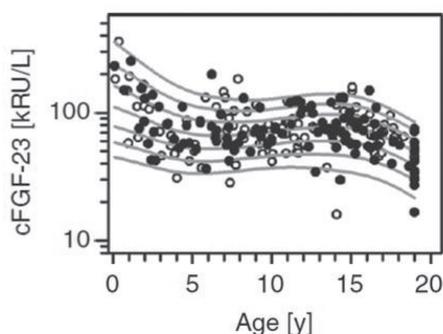
Sample preparation It is recommended to collect the sample in the morning after a 12-hour fasting period. Keep sample frozen at -20°C. Longer storage at -80°C. Avoid repeated freeze/thaw cycles.

Reference values

	Range (RU/ml)
Premenopausal women (<40 years)	20.9 – 91.1
Postmenopausal women (>60 years)	44.0 – 139.9
Men (27 – 76 years)	33.7 – 96.5

Children:

Group Age	n	Mean (RU/ml)	Range (RU/ml)
<1	8	105	43-324
2-4	27	80	36-197
5-11	73	69	35-132
12-15	61	77	36-138
16 - ≥19	49	61	27-107



Reference Fischer DC, Mischek A, Wolf A, Rahn A, Salveski B, Kundt G and Haffner D. Pediatric reference values for the C-terminal fragment of fibroblast-growth factor-23, sclerostin, bone-specific alkaline phosphatase, and isoform 5b of tartrate resistant acid phosphatase. *Annals of Clinical Biochemistry* 2012; 49: 546 -553.

Species Human, Cynomolgus Macaque

FGF-23 (C-Term) Mouse

FIBROBLAST GROWTH FACTOR 23

C-terminal

Cat. No.	60 - 6300
Tests	96
Method	ELISA
Range	30 – 1000 pg/ml (can be extended to 3000 pg/ml)
Sensitivity	4 pg/ml
Incubation time	3.5 hours
Sample volume	25 µl
Sample type	Serum, plasma, cell culture
Sample preparation	FGF-23 molecule is very unstable. Therefore samples should be assayed immediately or stored frozen at -20°C or below. Avoid repeated freeze/thaw cycles.
Species	Mouse, rat, canine, porcine

FGF-23 Intact, Human (Kainos)

FIBROBLAST GROWTH FACTOR 23

C-terminal

Cat. No.	CY-4000
Tests	96
Method	ELISA
Range	8 – 800 pg/ml
Sensitivity	3.0 pg/ml
Incubation time	3.5 hours
Sample volume	50 µl
Sample type	Serum / Heparin- and EDTA Plasma
Sample preparation	It is recommended to collect the sample in the morning after a 12-hour fasting period. Intact FGF-23 is very instable. Therefore, collection and testing or storage should take place promptly. Store samples at -20 °C or below. Avoid repeated freezing and thawing of specimens.
Reference values	10 – 50 pg/ml
Species	Human, Rat, Mouse

Klotho, Human

Cat. No.	27998
Tests	96
Method	ELISA
Range	93.75 - 6000 pg/mL
Sensitivity	6.15 pg/mL
Incubation time	2 hours
Sample volume	100 µL
Sample type	Serum, EDTA plasma
Sample preparation	Collection, testing and samples storage should take place promptly. Store samples at -20 °C or below. Avoid repeated freezing and thawing of specimens.
Reference values	239 – 1266 pg/mL
Species	Human
Specificity	No cross-reactions observed with: osteopontin, human VEGF or PDGF.

Intended use Klotho is a single-pass trans-membrane anti-aging protein (1014 amino-acids, 130 kDa, chromosomal location in 13q12 in humans) that has been recently shown to have wide and important biological effects. Its expression occurs mainly in the kidney and in the parathyroid. Its extra-cellular domain is wide whereas its intra-cellular part is very short; it also exists as a soluble protein.

In humans, Klotho polymorphisms have been associated both in a positive and negative way to bone mineral density, life expectancy, cardiovascular events (e.g., ischemic stroke, carotid atherosclerosis), biomarkers of metabolic syndrome (e.g., uric acid levels, lipid and glucose metabolisms), and even to cognitive ability.

TECOmedical Group

always your partner

Headquarters / Switzerland

TECOmedical AG

Gewerbestrasse 10
4450 Sissach

Phone +41 61 985 81 00

Fax +41 61 985 81 09

Mail info@tecomedical.com

Germany

TECOmedical GmbH

Wasserbreite 57
32257 Bünde

Phone +49 52 23 985 99 99

Fax +49 52 23 985 99 98

Mail info@tecomedical.com

Benelux

TECOmedical Benelux BV

Computerweg 22
3542 DR Utrecht, The Netherlands

Phone +31 30 307 87 30

Fax +31 30 870 49 39

Mail Benelux@tecomedical.com

Austria

TECOmedical AG

Phone 0800 20 40 66

Fax 0800 20 40 55

Mail info@tecomedical.com

www.tecomedical.com